

## Kuduz aşısının geliştirilmesi Pasteur tarafından geliştirilmesi

6 juillet 1880	1/2 iuv. moelle de 21 journ.	Morte 2. 15 jours
7 - 9 matin —	23 — morte de 14 journ.	
7 - 6 11/2 —	25 — moelle de 12 journ.	
8 - 9 matin —	27 — morte de 11 journ.	
8 - 6 6/2 —	29 ; — moelle de 9 journ.	
9 - 11 matin —	1er juillet moelle de 8 journ.	
10 — ~ — ~ —	3 — morte de 7 journ.	
11 — ~ — ~ —	5 — morte de 6 journ.	
12 — ~ — ~ —	7 — morte de 5 journ.	
13 — ~ — ~ —	9 — morte de 4 journ.	
14 — ~ — ~ —	11 — morte de 3 journ.	
15 — ~ — ~ —	13 — morte de 2 journ.	
16 — ~ — ~ —	15 — morte de 1 jour	

Image of Pasteur's handwritten table of the rabies vaccination procedure. English translation version of the table is below. See Fig. S1 for complete image of page. Image courtesy of Bibliothèque nationale de France.

## Half a syringe

		Marrow exposed to dry air on:	Marrow dehydration for:
6 July	8 pm	21 June	15 d*
7 July	9 am	23 June	14 d*
7 July	6 pm	25 June	12 d*
8 July	9 am	27 June	11 d*
8 July	6 pm	29 June	9 d*
9 July	11 am	1 July	8 d*
10 July	11 am	3 July	7 d*
11 July	11 am	5 July	6 d**
12 July	11 am	7 July	5 d**
13 July	11 am	9 July	4 d**
14 July	11 am	11 July	3 d**
15 July	11 am	13 July	2 d**
16 July	11 am	15 July	1 d**

\*Nonvirulent in rabbits.

\*\*Virulent in rabbits.

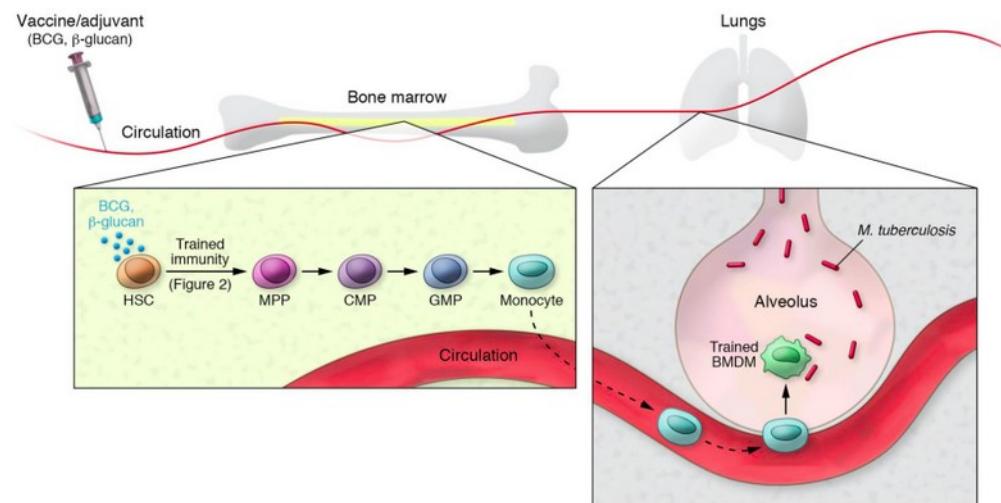
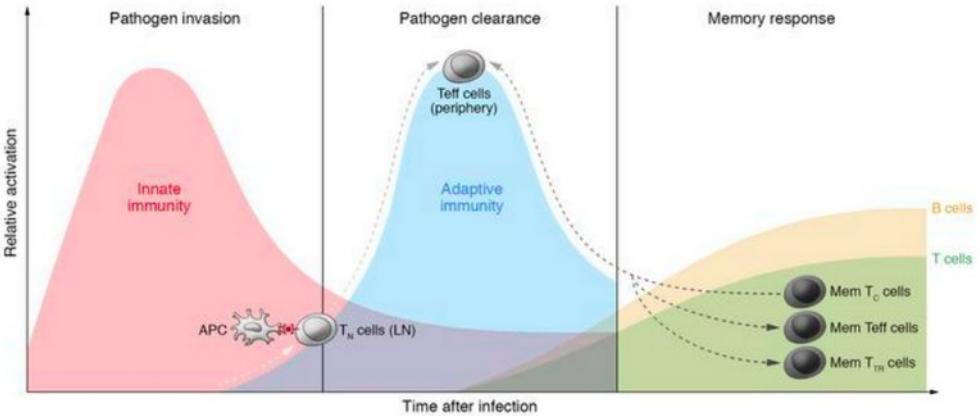


License: Public Domain Mark  
Credit: The Pasteur Institute, Kasauli, India: production of the rabies vaccine: caged rabbits showing symptoms of rabies after inoculation. Photograph, ca. 1910.



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Credit: The Pasteur  
Institute, Kasauli, India:  
production of the rabies  
vaccine: a rabbit, under  
anaesthetic, being  
infected with rabies by  
injection in the spine.  
Photograph, ca. 1910.



Khader SA, Divangahi M, Hanekom W, Hill PC, Maeurer M, Makar KW, Mayer-Barber KD, Mhlanga MM, Nemes E, Schlesinger LS, van Crevel R, Vankayalapati R, Xavier RJ, Netea MG; Bill and Melinda Gates Foundation Collaboration for TB Vaccine Discovery Innate Immunity Working Group18. Targeting innate immunity for tuberculosis vaccination. J Clin Invest. 2019 Sep 3;129(9):3482-3491. doi: 10.1172/JCI128877. PMID: 31478909; PMCID: PMC6715374.

Sepulveda-Crespo D, Resino S, Martinez I. Innate Immune Response against Hepatitis C Virus: Targets for Vaccine Adjuvants. *Vaccines (Basel)*. 2020 Jun 17;8(2):313. doi: 10.3390/vaccines8020313. PMID: 32560440; PMCID: PMC7350220.

PRR Type	Class of Activated Innate Receptor/Pathway	Adjuvant Name	Composition	Main Stimulated Immune Responses
TLRs	TLR3	Poly(I:C) or Poly(I:C) stabilized with poly-L-lysine	dsRNA analogs	Ab response, CD8 <sup>+</sup> T-cell response, Th1 type immunity
	TLR4	MPLA, GLA-SE, RC-529	MPL GLA AGP	Ab response CD8 <sup>+</sup> T-cell response Th1 type immunity
	TLR5	Flagellin fused to antigen	Bacterial flagellin	Ab response, Th1/Th2 response
	TLR7, 8 or both	Imiquimod/R837 (TLR7), Resiquimod/R848 (TLR7/8), 3M-052 (TLR7/8)	Imidazoquinoline analogs	Ab response, CD4 <sup>+</sup> /CD8 <sup>+</sup> T-cell response, Th1 type immunity
	TLR9	CpG-ODN	Synthetic ODN with optimized CpG motifs	Ab response, CD8 <sup>+</sup> T-cell response, Th1 type immunity
NLRs	RIG-I MDA-5	M8, Defective interfering RNA	dsRNA analogs	Ab response, CD4 <sup>+</sup> /CD8 <sup>+</sup> T-cell response
RLRs	Nod1 Nod2	iE-DAP, MDP	Bacterial peptidoglycan analogs	Ab response
CDs	STING	c-di-GAMP	Bacterial cyclic dinucleotides	Ab response, CD8 <sup>+</sup> T-cell response, Th1 type immunity

Ab: Antibody; AGP: Aminoalkyl glucosaminide 4-phosphate; c-di-GAMP: Cyclic guanosine monophosphate-adenosine monophosphate dinucleotide; CDs: Cytosolic DNA sensor ligands; CpG: Cytosine-phosphate-guanine; dsRNA: Double-stranded RNA; GLA: Glucopyranosyl lipid-adjuvant; iE-DAP:  $\gamma$ -D-glutamyl-meso-diaminopimelic acid; MDA-5: Melanoma differentiation-associated gene 5; MDP: Muramyl dipeptide; MPLA: Monophosphoryl lipid A adjuvant; NLRs: Nucleotide-binding oligomerization domain-like receptors; Nod1/2: Nucleotide oligomerization domain 1/domain 2; ODN: Oligodeoxynucleotide; Poly(I:C): Polyinosinic:polycytidylic acid; PRRs: Pattern recognition receptors; RIG-I: Retinoic acid-inducible gene-I; RLRs: RIG-I-like receptors; STING: Stimulator of interferon genes; Th1/Th2: T-helper cell type 1/type 2; TLRs: Toll-like receptors.

Lee BY, Mueller LE, Tilchin CG. A systems approach to vaccine decision making. Vaccine. 2017 Jan 20;35 Suppl 1(Suppl 1):A36-A42. doi: 10.1016/j.vaccine.2016.11.033. Epub 2016 Dec 22. PMID: 28017430; PMCID: PMC5460980.

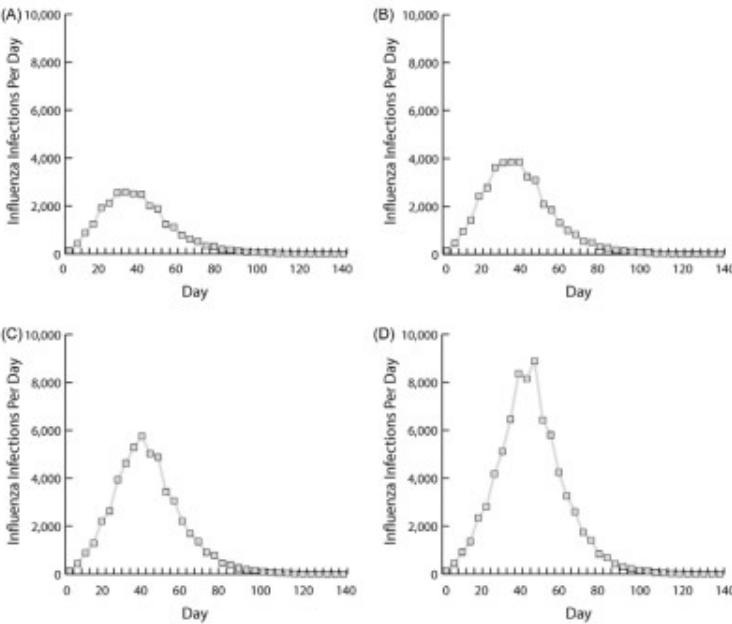
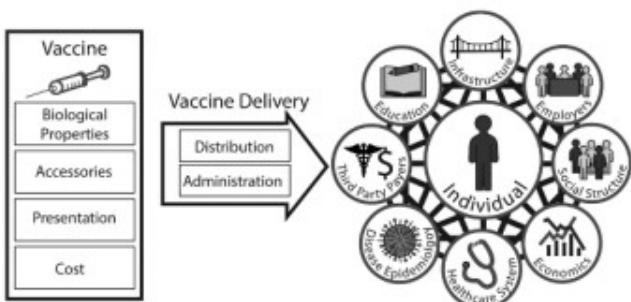
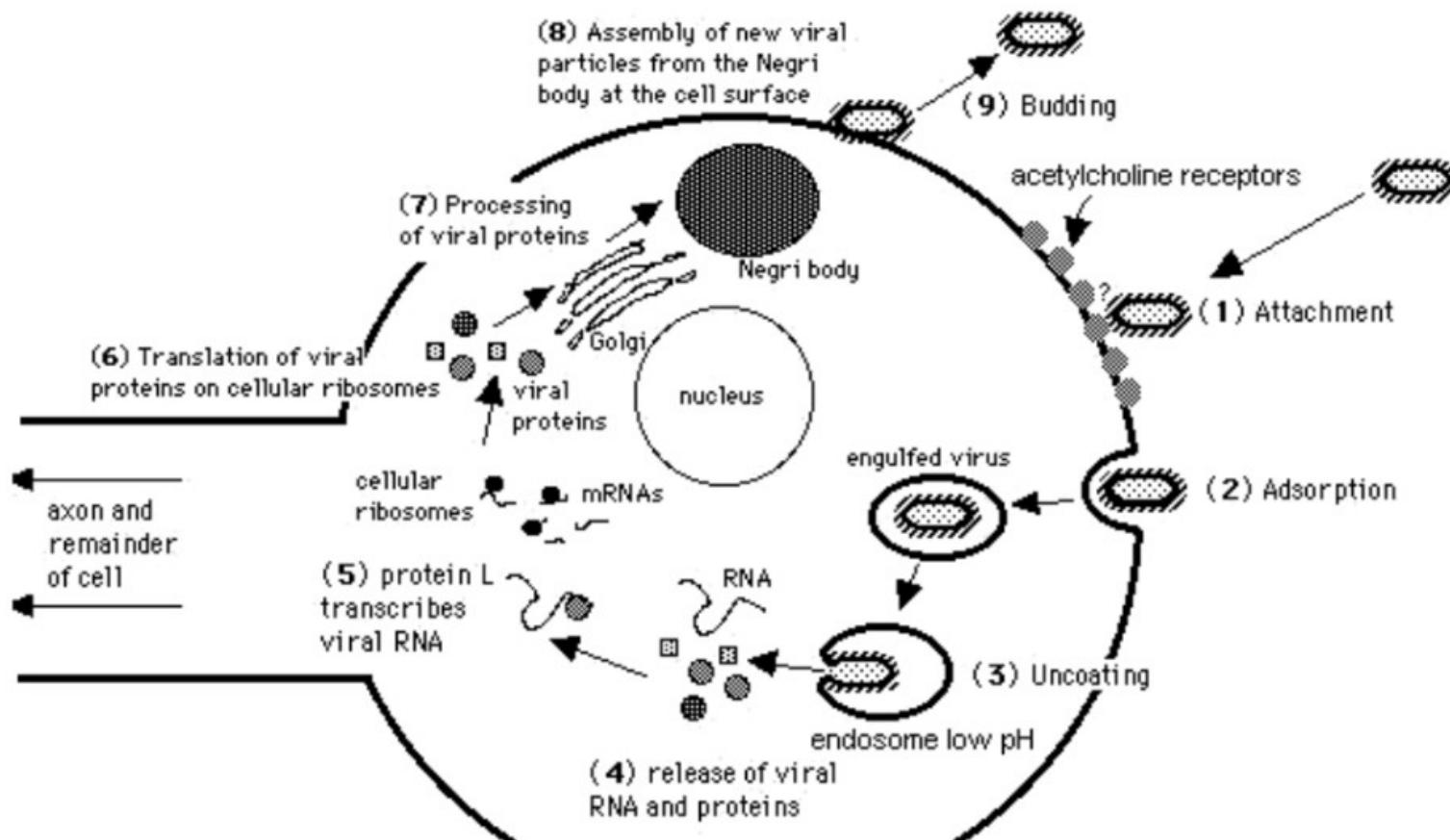


Fig. 5. Number of new influenza infections each day when two lowest-income counties experience vaccination delays of (a) 0 days, (b) 10 days, (c) 20 days, or (d) 30 days compared to other counties.

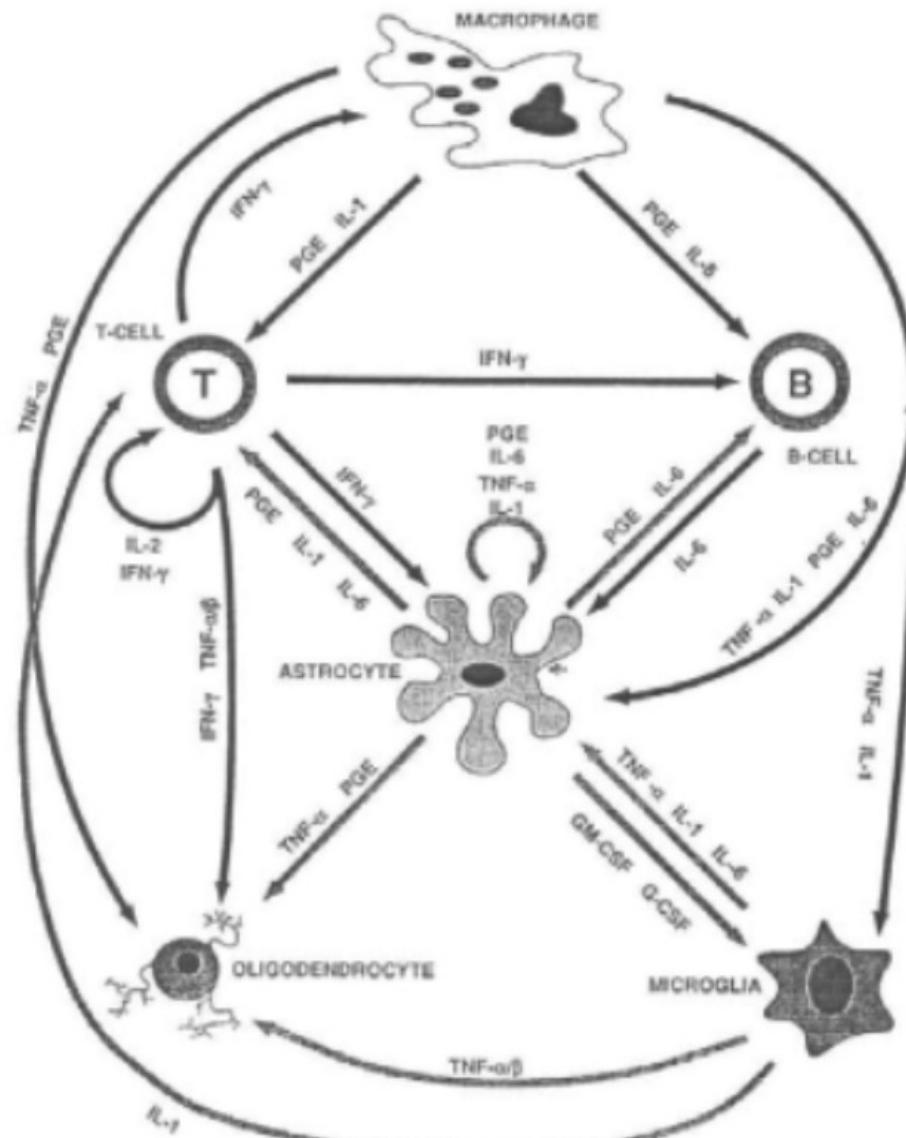
The mortality and severity of the target disease.  
The safety of the vaccine candidate.  
The full economic benefit of the vaccination program.

## Host Cell (Neuron) Infection



*J. Venom. Anim. Toxins incl. Trop. Dis.*  
V.13, n.1, p.5-38 , 2007.  
Review article.  
ISSN 1678-9199.

Figure 2: Cycle of viral infection and replication. Adapted from Mazaré



					Pneumothorax
					Intravascular
					thrombosis
					Secondary
					infections
					Pituitary dysfunction
					Hypoventilation, apnea
					Hypotension
					Cardiac arrhythmia, cardiac arrest
					Coma
					Hyperventilation, Hypoxia
					Aphasia, Incoordination.
					CNS signs; paresis, paralysis
					Hydrophobia, pharyngeal spasms
					Confusion, delirium, hallucinations
					Marked hyperactivity
					Anxiety, agitation, depression
					Fever
					Anorexia, nausea,
					Vomiting; headache
					Malaise, lethargy
	None				Pain or Paresthesias at bite site
					Death occurs
	Exposure	First	First neurological sign	Onset of	or recovery
		symptom		coma	begins
Clinical	Incubation	Prodrome	Acute neurological	Coma	Recovery
Stage	▼Period	▼	phase	▼	▼
Usual	20 to 90 days	2 to 10 days	2 to 7 days	0 to 14 days	Several months
Duration					

SPECIMEN	TEST	DETECTION
<b>ante mortem</b>		
skin punch biopsy; repeat until a diagnosis is obtained	FAT test on frozen section RT-PCR	antigen detection viral RNA virus isolation
saliva, tears, CSF; repeat until a diagnosis is obtained	Tissue culture Suckling mouse inoculation RT-PCR	virus isolation viral RNA
serum	Unvaccinated; test immediately* Vaccinated; save and compare a few days later	antibody detection
CSF	test immediately with serum*	antibody detection
<b>postmortem</b>		
brain needle necropsy** of two or more samples (brainstem and cerebellum)	FAT test of impressions smear RT-PCR suckling mouse inoculation	Antigen detection Viral RNA Viral Isolation
retrospective diagnosis	enzyme methods	antigen detection in formalin-fixed tissue

FAT: Fluorescent antibody test

RT-PCR: Reverse transcription polymerase chain reaction

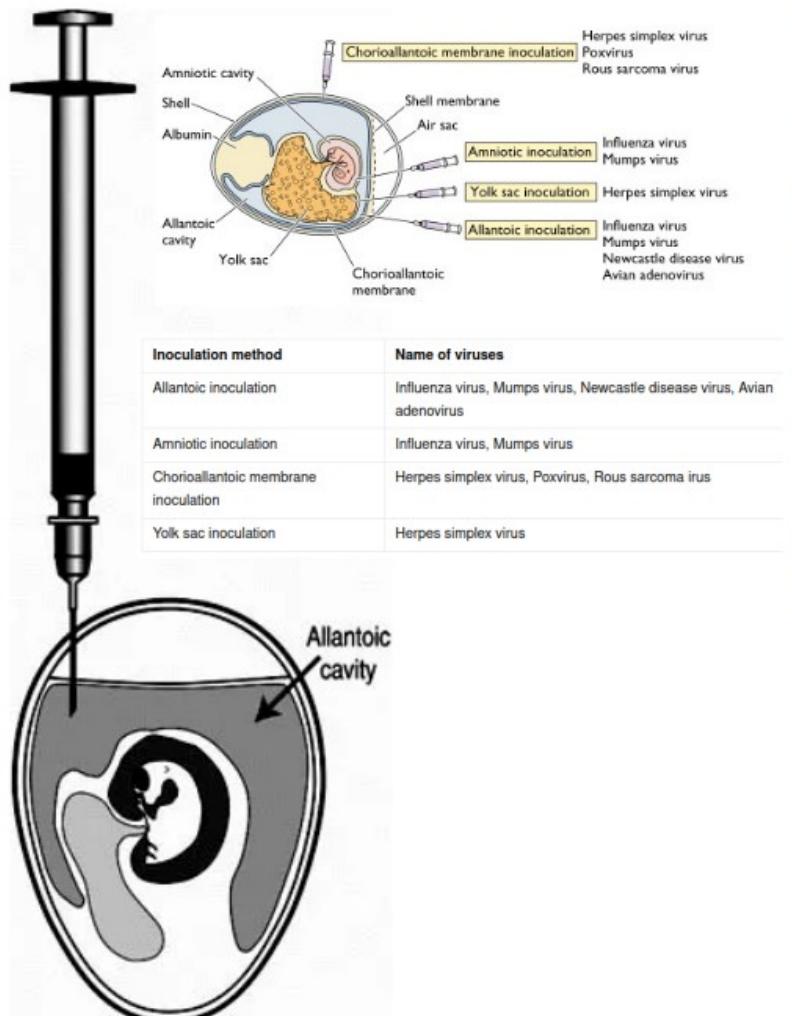
CSF: Cerebrospinal fluid

\* In unvaccinated patients, rabies antibody generally appears in the second week of the disease. Rabies specific IgM has been detected in the serum and in some cases in the CSF at low concentration, but no earlier than IgG. High concentrations of antibody in the CSF have been considered diagnostic despite vaccination.

\*\* Needle necropsy: with a Vim-Silverman's or other long biopsy needle.

Source: Warrell and Warrell, 2004 (94).

<https://www.virology.ws/2009/12/10/influenza-virus-growth-in-eggs/>



<https://microbeonline.com/embryonated-egg-cultures-viruses/>

