



# Akademik Yazım ve Sunum Becerileri

(Ders Notu\*)

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*\* Ders notunun hazırlanmasında kullanılan kaynaklar son sayfada toplu olarak verilmiştir.*

# Akademik Yazımda Temel Kurallar Nelerdir? (Akademik Dil Kullanımı)

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## Giriş

- Problem ve önemini açıklayın.
- Araştırma konusu ile alan ilişkisi nasıldır?
- Araştırmanın önemi açısından ilgili diğer araştırmalara değinin (tam bir literatür taraması şeklinde değil).
- Makalenin geri kalanını anlamak için gerekli tüm bilgileri verin.
- Kavramları açıklayın ve terimleri tanımlayın.
- Çalışmanın amacını açık ve net bir şekilde ortaya koyun.
- Tekrardan kaçının.
- Dergiye bağlı olmakla birlikte sınırlı harf/kelime

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## Giriş

### INTRODUCTION

There are many different practices such as tagging, tail docking, dehorning, castration, weaning, vaccination, bathing, hoof trimming and shearing in sheep husbandry. Some of these practices are performed only once in the animal's lifespan, while others are necessarily repeated periodically and these may cause an endocrine and metabolic response to occur in animals known as stress [1,2]. Animals that are exposed to stress react in species-specific behavioral patterns that are also influenced by learned behavior, and there are differences between animals in response to these behaviors. Animals, depending on their genetic structure and previous experience, can respond differently to the same stressors. Moreover age, gender, physiological status, herd density, daily rhythm and other environmental factors can also affect the individual reactions that animals show against the stressors [3,4]. Under different environment and management conditions, animals that are exposed

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## Giriş

### 1 Introduction

Insulin-like growth factors (IGFs) are one of the most important compounds that influence metabolism in animals. IGFs are a family of polypeptides that carry out metabolic and mitogenic activities. The most potent mitogenic IGF peptide is insulin-like growth factor-1 (IGF-1), a basic polypeptide of 70 amino acids directed by growth hormone under normal physiological conditions (Buonomo et al., 1988). IGF-1 is mainly produced in the liver in addition to being produced by the environmental tissues such as the skin, ovary, placenta, breast and bone as autocrine/paracrine (Hashizume et al., 2000; Basturk, 2007). The bioactivity of IGF-1 is achieved by specific insulin-like growth factor binding proteins (IGF-BPs) with high affinity (Obese et al., 2008).

IGF-1 plays an important role in various physiological processes such as reproduction, growth, lactation and the health of the organism (McGuire et al., 1992). IGF-1 is involved in the growth and function of almost every organ in the body (Rasouli et al., 2017). The predominant physiological effect of IGF-1 is the stimulation of postnatal body growth. In addition, IGF-1 can regulate the synthesis of whole body protein, the uptake of glucose by peripheral tissues and the regulation of lipid metabolism (Hadsell et al., 2002).

IGF-1 is secreted nonpulsatile, and the secretion level of IGF-1 is phenotypically correlated with the live weight and growth rate in cattle, pigs, sheep and chickens (Bishop et al., 1989). In addition, it has been reported that IGF-1 concentrations in farm animals are significantly affected by envi-

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## Materyal ve Metod

- Her biyolojik, analitik ya da istatistiksel işlem için, açık, tekrarlanabilir tanımlama sağlanmalıdır.
- Belirli bir orijinal referans, tanımlama yerine sunulabilir.
- Kullanılan metoddaki herhangi bir değişiklik yapıldıysa, bunun açıklanabilir ve haklı olması gerekmektedir.
- Kit kullanılan metodoloji için referans bildirin.
- Doğruluk ve hassasiyet, türe ve yaşa bağlı olarak değişebilir.

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## Materyal ve Metod (devam)

- Deneylerin doğruluğunu sağlayın.
- Tekrarlanan analizler için ortalama ve VK
- Hassasiyet (minimum miktar ve saptanabilir doz)
- Başkaları tarafından tekrarlanabilmesi için deneyin tüm detaylarını anlatın.
- Santrijüj devri, süresi ve sıcaklığı gibi
- Alınan kan hacmi, koruyucu veya antikoagülan miktarı (Heparin, EDTA, vb.)

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## **Materyal ve Metod (devam)**

- Rasyonu tanımlayın.
- Deneme aktivitelerinin tarihlerini bildirin (örneğin kan alım tarihleri)
- Denemede kullanılan hayvanların;
- Irk, cinsiyet, yaş, canlı ağırlık, fizyolojik durum, barındırma koşulları, yemleme yönetimi, vb. açıklayın.
- İnvaziv bir girişim varsa tekniğini açıklayın.
- Denemenin yapıldığı ünitenin koordinatlarını ve iklimsel verilerini açıklayın.

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## **Materyal ve Metod (devam)**

- Kullanılan istatistik yöntemi açıklayın.
- Eğer yöntem bu alanda yaygın kullanılıyorsa, referans vererek genel bir anlatım yapabilirsiniz.
- İstatistiksel modeli açıklayın.
- $P$  değerlerini verin.
- Deneme dizaynını açıklayın.



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## **Materyal ve Metod (devam)**

Bu kısımda;

- **Sadece gerekli olan bilgiler verilmelidir.**
- Bulgular verilmez.
- Sonuçlar verilmez.
- Tartışma yapılmaz.
- Yorum yapılmaz.

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## Materyal ve Metod (devam)

### MATERIALS AND METHODS

#### Animal care

The experimental procedure was approved by Local Ethics Committee at Erciyes University (16/004).

#### Location, experimental animals, shearing procedures, and experimental groups

This study was carried out between June and September months of 2016 in a farm entitled Erciyes University Agricultural Research and Application Centre (ERUTAM) located in Kayseri, Turkey (38°29'21.5" N; 35°10'11.7" E) at an altitude of approximately 1,130 meters above sea level. The study was conducted on 1,5 years old 39 non-pregnant Akkaraman ewes at the beginning of the experiment. The 39 ewes were chosen randomly from the flock belonging to the Erciyes University. During 15 days prior to starting of the study, all ewes were identified and subjected to internal and external parasite controls. The following experimental protocol was used: the ewes were randomly divided into two groups; i) Group A (n = 20) designed as the control group, and were shorn and group B (n = 19) designed as the experimental group, and were unshorn. The shearing was performed via sheep shearing machine (Xpert 708-200, Heiniger AG, Herzogenbuchsee, Switzerland) 11:00 am and 2:00 pm on the same day, without any interruption. All the ewes were clinically healthy. During the experimental period, the ewes grazed

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## Materyal ve Metod (devam)

### 2 Materials and methods

#### 2.1 Experimental animals, location and management

This study was carried out on 13 White goat (75 % Saanen and 25 % Kilis goat) kids and 12 Angora goat kids raised in the Animal Husbandry Station (39°57'42.5" N, 32°51'56.2" E) at the Ankara University, Faculty of Agriculture, Department of Animal Science. All of the goat kids were clinically healthy. The animals were housed in shaded pens with their dams, and natural light from windows and a door was allowed to pass through to the pens. The goat kids were allowed to receive milk from their dams during the experimental period, and from 2 weeks old they were they were fed concentrate feed and alfalfa hay ad libitum. Experimental feeds were collected during the experimental period and analyzed for chemical composition. Dry matter, crude protein, crude fiber, crude ash and crude fat in feeds were analyzed according to the AOAC Official Methods of Analysis (AOAC, 2000). Neutral detergent fibre (NDF) and acid detergent fibre (ADF) were measured according to the methods described by Van Soest et al. (1991). The metabolizable energy (ME) of forage and concentrate mix was calculated according to the Turkish Standards Institute (TSE, 1991). The concentrated feed composition and ME was as follows: dry matter (DM) comprised 91 %, crude protein (CP) comprised 18.05 %, crude fiber (CF) comprised 8 %, crude ash comprised 7.5 % and ether extract (EE) comprised 3.5 %; the ME was 2615 Kcal kg<sup>-1</sup>. The alfalfa composition and ME was as follows: DM comprised 91.5 %, CP comprised 13.0 %, CF comprised 34.0 %, crude ash comprised 8.29 %, EE comprised 1.00 %, ADF comprised 8.0 %, NDF com-

prised 49.0 %; the ME was 2000.22 kcal kg<sup>-1</sup>. Fresh water was always available to the goat kids. Management of experimental goat kids did not interfere with the general operation of the station, and the study was conducted within standard ethical norms. Some properties of experimental goat kids are shown in Table 1.

#### 2.2 Blood collection and IGF-1 analysis

During the experimental period, the first samples (Period 1) were taken 14–15 d after birth, and samples (periods 2–10) were taken at 15 d intervals for 5 months. On sampling days, blood samples were regularly taken from the vena jugularis of each goat kid using vacuum containers without anticoagulant (VACUETTE® TUBE 8 mL Z Serum Sep Clot Activator). The blood samples were centrifuged at 4000 × g for 5 min, and the serum was stored at –20 °C until the analysis was carried out. IGF-1 concentrations were determined in the blood serum using a commercial ELISA kit (Fine Test Goat IGF-1 Cat. no. EG0002). The intra- and inter-assay coefficients of variation were < 8 % and < 10 %, respectively. The least detectable concentration was 14 ng mL<sup>-1</sup>.

#### 2.3 Measurement of body traits

With respect to body trait measurements, the body weight (BW), withers height (WH), rump height (RH), body length (BL), chest depth (CD), chest width (CW), chest girth (CG) and cannon bone circumference (CBC) of each kid were regularly measured on the same sampling days. BW was measured using a commercial hanging scale (±10 g). A measuring tape was used to measure the height, length, depth and width of the goat kids, whereas other body measurements were taken using a flexible tape measure. All measurements were taken by the same operator and followed the methodology of Herrera et al. (1996).

# Kaynakça

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